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LC-MS-MS Analysis of Terfenadine and Its Metabolites in Rat Plasma Using Oasis[®] MCX

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he high performance liquid chromatography (HPLC) system used was a Waters Alliance 2795 HPLC system equipped with a sample chiller and an in-line solvent degasser. The analytical column was an XTerra MS C18 column (2.1 mm × 30 mm, 3.5 μm). AMic romass Quattro Ultima triple quadrupole mass spectrometer was equipped with an electrogray source and set in positive acquisition using multiple reaction monitoring (MRM). Data acquisition and instrument control we re powered by MassLynxTM chromatographic software (Mic romass, Ltd., United Kingdom). The μElution SPE 96-well plate was packed with 2 mg of Oasis MCX.

Experimental Conditions

Sample preparation: Pipette 5 mL of raw uncentrifuged rat plasma (heparin as anticoagulant) into ten 10 mL scintillation vials for the calibration curve. Add mixture of terfenadine, terfenadine–alcool, and terfenadine–carboxylate at various concentrations to achieve a calibration from 0.5 ng/mL to 200 ng/mL. Add 100 μL of concentrated phosphoric acid to all vials. Condition the Oasis MCX with 200 μL of methanol and pull through with a positive or negative displacement. Repeat with 200 μL of water. Load 250 μL of spiked rat plasma in each well, followed by 250 μL of internal standard (protriptyline) in water at 10 ng/mL. Apply vacuum to dryness. Wash with 200 μL of water and 0.1 N HCl, followed by 200 μL of methanol. Elute with 25 μL of 40:60 acetonitrile–isopropanol and 5% ammonium hydroxide in a 350-μL or 1-mL collection plate. Dilute the eluent with 50 μL of water and inject 25 μL of the resulting solution onto the LC system.

Table I: Analysis of terfenadine and its metabolite in rat plasma using Oasis MCX				
	Concentration (ng/mL)			CV (%)
Terfenadine	0.5	0.51	0.006	2.0
	1.0	0.96	0.044	4.6
	5.0	4.99	0.240	4.8
	10.0	10.11	0.46	1.8
	20.0	19.26	0.355	1.8
	50.0	54.05	1.96	3.6
	100.00	103.25	3.36	3.2
	200.00	197.18	1.82	0.9
Terfenadine–alcool	0.5	0.494	0.010	2.3
	2.0	1.935	0.082	4.2
	5.0	5.05	0.246	4.8
	10.0	10.12	0.454	4.4
	25.0	24.82	1.157	4.6
	50.0	50.51	2.22	4.3
	100.0	101.69	3.03	2.9
	200.0	196.39	4.12	2.1
Terfenadine–carboxylat	ite 0.5	0.51	0.0075	1.4
	1.0	0.975	0.038	3.9
	5.0	5.192	0.254	4.9
	10.0	10.41	0.565	35.4
	20.0	19.80	0.722	3.6
	25.0	24.66	0.556	2.2
	50.0	51.08	1.72	3.3
	100.0	102.05	2.95	2.8
	200.0	196.55	7.40	3.7

Chromatographic conditions: The analytical column was an XTerra MS C18 column (2.1 mm \times 30 mm, 3.5 μ m) used with a gradient mobile phase consisting of 0.1 M ammonium formate at pH 9.5 in water (line A) and methanol (line B). Four compounds we re separated in 4 min using gradient conditions at 0.4 mL/min with an injection volume of 25 μ L. The gradient starts at 5% organic and ramps at 95% organic in 1 min, stays at 95% organic for 1 min more, drops to the original condition in 30 s, and reconditions for 2.5 min. The QuattroUltima triple quadrupole system was set for analysis with electrospray in positive mode using MRM. The quantitation was performed on the MRM transition 472.2 \rightarrow 436.3 for terfenadine, 502.2 \rightarrow 466.2 for terfenadine–carboxylate, 488.2 \rightarrow 452.2 for terfenadine–alcool and 263.9 \rightarrow 190.8 for the internal standard (protriptyline).

Results and Conclusion

The average recove ry of terfenadine and its metabolite (n=6), shown in Table I, was 85% and higher with an average coefficient of variation (CV) of 5%. In this situation, sub-nanograms per milliliter LOQ was reached with a simple mixed-mode generic method and gave higher sensitivity. The two-step cleanup in the Oasis MCX protocol removes more interferences than the generic Oasis HLB.

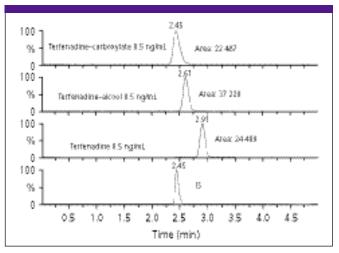


Figure 1: Analysis of terfenadine in rat plasma using Oasis MCX.

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